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Section: Education Manual	Subject Title: Mycology - Training Manual for Residents	
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# **MYCOLOGY - TRAINING MANUAL FOR RESIDENTS**

### Teaching and Supervising Technologist: Subhash K. Mohan

Reference:

- Davise H. Larone: Medically Important Fungi, A guide to identification, 3<sup>rd</sup>. Edition, ASM Press, 1995
- 2. Guy St Germain, Richard Summerbell; Identifying Filamentous Fungi, A Clinical Laboratory Handbook, Star Publisher, 1996
- 3. Martha E Kern: Medical Mycology; Self Instructional Text, F A Davis, 1985 (1997)
- 4. Selected Reference Articles

## The Trainee at all levels of training shall know:

### 1.0 *Processing of Specimens*

- 1.1 Know about proper mycology specimen collection, transportation and planting and the use of selective and/ or non-selective media
- 1.2 State the appropriate incubation temperature and length of incubation for recovery of various pathogens

### 2.0 Direct Microscopy

- 2.1 Demonstrate the proper of use of microscopy in the detection of fungal elements in clinical specimens using: Fungi-Fluor (FS), 10% KOH and Calcofluor White (CW)
- 2.2 Demonstrate and interpret fungal elements examined microscopically such as yeast, pseudo hyphae, septate or non-septate hyphae, narrow or broad base attachment etc. and/ or suspect or identify certain pathogens presumptively based on structure
- 2.3 Explain the different uses, advantages and disadvantages of the FS, CW and 10% KOH

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#### 3.0 **Procedure for identification of Yeast**

- 3.1 Demonstrate the ability to identify various yeasts using Germ Tube, Oxgall, Cornmeal, Urea, EBM, Rapid Yeast ID System, API 20C procedures
- 3.2 Explain the principles, uses, advantages and disadvantages of the above methods
- 3.3 Ability to recognize organisms looking like yeast such as Prototheca based on structure

### 4.0 *Protocol for reporting results as per manual policy*

- 4.1 Reporting positive FS results in SOFT and phoning critical results to ward
- 4.2 Reporting positive culture results in SOFT and phoning ward where applicable

### 5.0 *Procedures for identification of moulds*

- 5.1 Demonstrate the following methods used: Macroscopic Examination of Cultures, Microscopic Examination of Cultures, Scotch Tape Preparation, Tease Mount Preparation, Slide Culture, Sub-Culture, Phase Conversion, Permanent Mounts and Stock Preparation
- 5.2 Explain the purpose of each of the above

### 6.0 *Identification of the following fungi to the genus and/ or species level where indicated:*

#### Rapid Grower:

Aspergillus fumigatus, niger, flavus and terreus Penicillium species and Paecilomyces species Fusarium species and Acremonium species Scopulariopsis species and Trichoderma species Scedosporium species (apiospermum and prolificans) and P. boydii

#### Zygomycetes:

Rhizopus species Mucor species Absidia species

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### Dematiaceous Moulds:

Cladosporium species Curvularia species and Bipolaris species Alternaria species and Exophiala species

#### Superficial Mycotic Agents:

Dermatophytes and Scytalidium species Trichosporon beigelii Malassezia furfur (Pityriasis versicolor – microscopic only, no culture necessary)

### Dimorphic Fungi:

Histoplasma capsulatum and Blastomyces dermatitidis Sporothrix schenckii and Penicillium marneffei (C. immitis – Demonstration only)

### 7.0 Identification of Actinomycetes and other Filamentous Organisms

Nocardia species Streptomyces species (Compare to Rapid Grower Mycobacteria)